

“Pharmacokinetics and efficacy of meloxicam  
administered subcutaneously and intramuscularly  
in sheep”

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## 1 **Abstract**

2 **Objective** To determine the pharmacokinetic profiles and efficacy of meloxicam when  
3 administered subcutaneously (S/C) and intramuscularly (IM) in sheep at different doses.

4 **Procedures** Ewes were injected with 0.1mL of oil of turpentine in a forelimb, followed by  
5 either a 1.0mg/kg or 2.0mg/kg dose of meloxicam administered either subcutaneously or  
6 intramuscularly (n=3). Ewes were examined at 0, 0.5, 1, 2, 4, 6, 8, 10, 12, 24 and 48 h, with  
7 blood collected (3-4mL) at each timepoint and behavioural and physiological responses  
8 recorded. Responses measured included skin temperature, limb circumference, limb  
9 sensitivity and gait. Pharmacokinetics were analysed using HPLC analysis.

10 **Results** Oil of turpentine successfully induced inflammation in the ewes with affected limbs  
11 demonstrating higher skin temperature ( $p < 0.001$ ), limb circumference ( $p < 0.001$ ) and  
12 sensitivity ( $p = 0.01$ ). Whilst significantly affected by meloxicam, minimal variations of skin  
13 temperature ( $p < 0.001$ ), limb circumference ( $p = 0.012$ ) and gait ( $p < 0.001$ ) were observed  
14 between treatments. S/C 1.0mg/kg, IM 2.0mg/kg and S/C 2.0mg/kg treatments significantly  
15 reduced limb sensitivity when compared to the control at 48hrs post drug administration ( $p$   
16  $< 0.001$ ). The IM treatment at 1.0mg/kg had a significantly higher plasma concentration of  
17 meloxicam than S/C at 1.0mg/kg from 0.5-4hrs post drug administration ( $p < 0.001$ ). Both IM  
18 and SC treatments demonstrated long terminal half-lives at 12.47 and 10.24 hrs respectively.

19 **Conclusion** Meloxicam was effective at providing some analgesia post-injection of turpentine  
20 in sheep however analgesic efficacy could not be distinguished between the five treatments  
21 (IM 1.0mg/kg, S/C 1.0mg/kg, IM 2.0mg/kg, S/C 2.0mg/kg and control).

22 **Key words**

23 analgesia; animal welfare; meloxicam; pharmacokinetics; sheep

24 **Abbreviations**

25 AUC = Area under the curve

26 C<sub>max</sub> = Maximum serum concentration

27 T<sub>1/2</sub> = Half-life of the terminal portion of the curve

28 T<sub>max</sub> = Time to maximum serum concentration

29 COX = cyclo-oxygenase

30 NSAID = non-steroidal anti-inflammatory drug

31 **Introduction**

32 Meloxicam is a non-steroidal, anti-inflammatory drug (NSAID) currently registered for pain  
33 management in a number of species including humans, cats, dogs,<sup>1</sup> cattle, pigs and most  
34 recently sheep.<sup>2</sup> Currently meloxicam is the only registered NSAID for use in sheep,<sup>3</sup> with  
35 Metacam<sup>®</sup> 20 (Boehringer Ingelheim, Australia) officially registered for use in 2016.<sup>2</sup>  
36 Meloxicam is an enolic acid that provides analgesia and antipyretic action via the inhibition of  
37 the cyclo-oxygenase (COX) pathway.<sup>4</sup> The COX pathway is responsible for the biosynthesis of  
38 arachidonic acid to prostaglandins (PGE<sub>2</sub>), which impart pain via the stimulation of peripheral  
39 sensory neurons.<sup>5</sup> Meloxicam selectively inhibits the COX-2 pathway in a number of species  
40 including humans,<sup>6</sup> primates<sup>6</sup> and horses,<sup>7</sup> resulting in the low ulcerogenic potential of the  
41 drug, alongside other favourable characteristics for animal use, including an extended  
42 elimination half-life and effective bio-absorption.<sup>4</sup> Meloxicam has been shown to significantly

43 reduce abnormal behaviours associated with pain during husbandry procedures such as  
44 castration and tail docking in sheep.<sup>8</sup>

45 The pharmacokinetics of meloxicam have been documented in numerous species including  
46 goats,<sup>9</sup> horses,<sup>10</sup> cattle<sup>11</sup> and sheep.<sup>1</sup> However, the pharmacokinetics of meloxicam in sheep  
47 have only been assessed via intravenous and oral routes of delivery with no literature  
48 currently available in regards to subcutaneous and intramuscular administration routes.<sup>1</sup>  
49 Metacam® 20 administration guidelines in sheep recommend a subcutaneous injection of  
50 1.0mg/kg,<sup>2</sup> therefore it is surprising that a knowledge gap is present in regards to the  
51 pharmacokinetics and efficacy of subcutaneous administration in sheep at the recommended  
52 dose. With known benefits such as on-farm practicality and slower absorption resulting in the  
53 potential for longer-lasting analgesia,<sup>12</sup> subcutaneous administration is ideal, however with a  
54 lack of knowledge available in sheep, alternative routes of delivery, such as intramuscular,  
55 may prove to be more efficacious for providing pain relief. Whilst there is no literature  
56 currently available on pharmacokinetics of intramuscular administration of meloxicam in  
57 sheep, previous studies investigating alternative NSAID's in pigs, sheep and horses indicate  
58 greater bioavailability and maximum concentrations when utilising intramuscular routes  
59 compared to alternative routes investigated.<sup>13-15</sup>

60 The objective of this study was to determine the pharmacokinetic profiles and efficacy of  
61 meloxicam when administered subcutaneously and intramuscularly in sheep at 1.0mg/kg and  
62 2.0mg/kg for the alleviation of pain and inflammation in sheep.

## 63 **Methods**

### 64 ***Sheep***

65 The experiment was conducted at a University of Sydney farm (“Mayfarm”), in Camden, New  
66 South Wales (NSW) and was approved by the University Animal Ethics Committee. 14 Merino  
67 ewes (44-68kg) were housed in a group pen (10x10m) underneath a covered shed with  
68 outdoor access. Two weeks prior to the commencement of the experiment, sheep were  
69 housed in the group pen in order to acclimatise to the experimental environment. Sheep had  
70 their necks shaved to expose the jugular region for precise jugular venepuncture during  
71 experimentation. During this time, sheep were also drenched for internal parasites with Q  
72 drench (Jurox, Hunter Valley, NSW) at the dose recommended by the manufacturer. 1 week  
73 prior to experimentation, sheep were habituated to experimental conditions (once daily  
74 5d/wk). This included catching, tipping and restraining each individual sheep in lateral  
75 recumbency and recurrent jugular venepuncture. Sheep were fed an allocation of 750g/d  
76 lucerne hay cubes (MultiCube®; 18% crude protein dry matter; 9.1 MJ/kg dry matter). During  
77 the 1-week washout period sheep were released onto a 3 ha paddock with ad libitum access  
78 to Kikuyu pasture and water.

### 79 ***Drug administration***

80 A randomised crossover design with a 10-day washout period was used, which is adequate  
81 for drug clearance in sheep.<sup>1</sup> 5 days prior to experimentation, sheep were weighed to  
82 calculate drug doses of Metacam 20® (Meloxicam 20mg/mL, Boehringer Ingelheim, Australia).  
83 Inflammation was induced via the injection of oil of turpentine (Sigma-Aldrich) as previously  
84 utilised by Colditz et al. (2011). Oil of turpentine (0.1mL) was injected subcutaneously via an  
85 18-gauge needle on the pastern midway between the fetlock and the coronet on a single  
86 forelimb of sheep. The limb chosen for turpentine injection was randomised across individual

87 sheep, alternating between the left and right forelimb, with the unaffected limb acting as a  
88 control.

89 Additionally, sheep were assigned to one of five treatment groups; 1.0mg/kg meloxicam  
90 (Metacam 20®; Boehringer Ingelheim) administered intramuscularly (n=3), 1.0mg/kg  
91 administered subcutaneously (n=3), 2.0mg/kg administered intramuscularly (n=3), 2.0mg/kg  
92 administered subcutaneously (n=3) or a control group (n=2) administered no dose of  
93 meloxicam (Table 1). Treatments were randomly assigned using a random number generator.  
94 Administration was undertaken using a 6-mL syringe and therefore doses were rounded to  
95 the nearest tenth in order to ensure precise meloxicam administration.

#### 96 ***Blood collection***

97 Blood samples were collected for pharmacokinetic analysis of meloxicam when administered  
98 at alternative routes and doses. Sheep were restrained in an upright position and blood  
99 samples (3-4mL) were collected into lithium heparin vacutainers via jugular venepuncture at  
100 0, 0.5, 1, 2, 4, 6, 8, 10, 12, 24 and 48 hours post drug administration. Samples were  
101 immediately centrifuged for 10 minutes at 3,500 X g. Plasma was transferred to sterile vials  
102 and frozen at -70°C until analysis. Analysis of samples took place within 35 days of collection.

#### 103 ***Skin temperature***

104 Skin temperature was measured using an infrared thermometer (non-contact thermometer,  
105 Jaycar Electronics) with a resolution of 0.1°C. The laser light on the thermometer was aimed  
106 at the cranial surface of both affected and unaffected carpal joints, in the region of the  
107 scaphoid and lunate bones, and held at the recommended distance (300mm). The same  
108 investigator was responsible for all skin temperature measurements to minimise inter-  
109 observer variation. Ambient temperature was also recorded for each collection time-point

110 (USB Temperature/humidity datalogger with LCD, Jaycar Electronics). Measurements were  
111 taken at 0, 0.5, 1, 2, 4, 6, 8, 10, 12, 24 and 48 hours post drug administration.

### 112 ***Limb circumference***

113 Limb circumference was measured, to the nearest millimetre, around the proximal aspect of  
114 the carpus by use of anatomic reference points, on both the affected and control limb using  
115 a measuring tape. The same investigator was responsible for all limb circumference  
116 measurements to minimise inter-observer variation. Measurements were taken at 0, 0.5, 1,  
117 2, 4, 6, 8, 10, 12, 24 and 48 hours post drug administration.

### 118 ***Limb sensitivity***

119 Sensitivity of the affected and unaffected forelimb was measured with a calibrated hand-held  
120 pressure algometer (Wagner Pain Test FPIX Digital Algometer, Wagner Instruments, Riverside,  
121 CT, USA) which has a maximum pressure of 10kg/f. The device consisted of a 1cm<sup>2</sup> blunt  
122 rubber tip and was applied, with increasing pressure at a perpendicular angle to the target  
123 site, midway between the fetlock and the coronet, on both the affected and unaffected limb  
124 of the animal. The force required for withdrawal of the limb was recorded as the mechanical  
125 nociceptive threshold (MNT) to the nearest 0.5kg/f. Sheep that were unresponsive to the  
126 applied force were recorded at the maximum threshold of 10kg/f. The hand-held device was  
127 returned to zero after each pressure test. The MNT was recorded in the second phase of  
128 experimentation by a single investigator to minimise inter-observer variation. Measurements  
129 were taken at 0, 0.5, 1, 2, 4, 6, 8, 10, 12, 24 and 48 hours post drug administration.

### 130 ***Gait score***

131 Sheep were assessed for gait using a numerical rating scale (NRS) based on behavioural  
132 characteristics of lameness (Table 2). Scoring took place once sheep were released back into  
133 the group pen after final measurements were taken for physiological and behavioural

134 indicators of pain. The observer stood within the group pen at a distance of 1-2 metres from  
135 the flock, observing voluntary movement in individual sheep for 1-3 minutes in order to  
136 obtain a lameness score. If individual sheep were out of sight, the observer entered the flight  
137 zone in order to stimulate movement within the flock and allow view of the desired individual.  
138 Measurements took place at 0, 0.5, 1, 2, 4, 6, 8, 10, 12, 24 and 48 hours post-injection by a  
139 single observer to minimise inter-observer variation.

#### 140 ***Serum meloxicam concentration measurement***

141 The concentration of meloxicam in serum was measured using high-pressure liquid  
142 chromatography (HPLC) analysis. The methodology was previously developed and validated  
143 for the University of Sydney HPLC laboratory in which analysis took place.<sup>16</sup> The HPLC system  
144 was comprised of a Shimadzu CBM-20A module (Kyoto, Japan) equipped with a LC-20AT  
145 delivery unit with DGU-20As degassing solvent delivery unit and SIL-20AC auto injector.

146 A reversed phase C<sup>18</sup> column (Synergi<sup>TM</sup> 4 $\mu$ m MAX-RP 80A, 150 x 4.6mm, Phenomenex, Lane  
147 Cove, NSW) was used for separation. The isocratic mobile phase comprised of 50mM  
148 potassium phosphate buffer (pH 2.15) and acetonitrile (55:45, v/v). The mobile phase was run  
149 at a flow rate of 1 mL min<sup>-1</sup> with an oven temperature of 30°C. The solution was monitored at  
150 a wavelength of 355nm via an SPD-M20A diode array detector (Kyoto, Japan) and Shimadzu  
151 class VP data system (software version 7.4) (Kyoto, Japan). The HPLC method was validated  
152 prior to the analysis of samples.

153 Meloxicam extraction was undertaken by the addition of 400 $\mu$ L of acetonitrile, containing the  
154 internal standard (IS) piroxicam, to 200 $\mu$ L of the serum sample (2:1 ratio). A vortex was used  
155 for 5 seconds to mix the samples and centrifugation at 14,000 g resulted in a precipitate.  
156 100  $\mu$ L of supernatant was pipetted to an injection vial, with the HPLC injection volume set to



157 20µL. Meloxicam plasma concentration within the samples was defined by obtaining standard  
158 curves via the analysis of blank plasma samples which were spiked with meloxicam.  
159 Meloxicam was observed at 8.5 minutes and the IS at 5.5 minutes (Figure 1.)

160 The standard curve used for determining meloxicam concentration through use of sheep  
161 plasma was linear from 0.049 to 25µg/mL. The lowest limit of quantification (LLOQ) was  
162 determined using the formula  $LLOQ = 10 \times \sigma/S$  whereby  $\sigma$  refers to the standard deviation  
163 (SD) and S refers to the slope of the calibration curves<sup>16</sup>. The LLOQ was defined as 0.096 µg/mL  
164 with the acceptance threshold defined as precision less than 15% and accuracy within  $\pm 20\%$   
165 of the nominal concentration across analyses as recommended by the International  
166 Guidelines for Bioanalytical Method Validation.<sup>17</sup> Intra-day accuracy as determined by the  
167 formulae:  $[(\text{estimated value}/\text{nominal value}) \times 100]$ <sup>16</sup> was  $102 \pm 2.96\%$ . Intra-day precision as  
168 calculated using the formula: coefficient of variation (CV)  $\times [(\text{SD}/\text{mean value}) \times 100]$ <sup>16</sup> was  
169  $0.97 \pm 0.87\%$ . Intra-day accuracy and precision were determined through triplicates for  
170 meloxicam concentrations of 0.2, 2.0, and 20.0 µg/mL across 3 consecutive days.

### 171 ***Pharmacokinetic analysis***

172 As a result of time constraints only S/C and IM 1.0mg/kg treatments were analysed using  
173 HPLC. The PK parameters calculated were: area under plasma concentration vs. time curves  
174 AUC<sub>0-t</sub>; where t refers to the final data time-point, maximum concentration of meloxicam in  
175 plasma (C<sub>max</sub>), time taken to reach maximum concentration in plasma (T<sub>max</sub>) and terminal  
176 half-life ( $t^{1/2}$ ). AUC<sub>0-t</sub> was calculated using the linear trapezoidal rule for approximation of the  
177 definite integral. C<sub>max</sub> was calculated by obtaining  $\bar{x}$  of the maximum concentrations of the  
178 samples (n=3). T<sub>max</sub> was calculated by obtaining the  $\bar{x}$  of the time taken to reach maximum

179 concentration for the samples where  $\bar{x}$  is the mean. The formula used for  $t_{1/2} = 0.693/kel$   
180 where kel is the elimination rate constant.

### 181 ***Statistical analysis***

182 Skin temperature, limb circumference and limb sensitivity were analysed in GenStat (VSN  
183 International Ltd, 14<sup>th</sup> Edition 2011) with a restricted maximum likelihood linear mixed  
184 model (REML). The model fitted the effects of treatment, timepoint and limb with a random  
185 effect of sheep ID. Ambient temperature was statistically significant against skin  
186 temperature and was therefore included in the model for skin temperature as a random  
187 effect. Limb sensitivity data was converted to binomials; 10kg/f = 1 and <10kg/f = 0 for  
188 statistical analysis. This was undertaken as transformation did not assist in normalising the  
189 dataset. Gait scores were subjected to ordinal logistic regression (OLR) in ASReml® 3.0  
190 statistical software (VSN International, Hemel Hempstead UK). The fixed effects of the  
191 model were treatment x timepoint and the random effect of the model was sheep ID. Data  
192 from the OLR analysis are presented as a cumulative odds ratios with the statistical  
193 probabilities of sheep having gait scores of  $Y = 0, 1, 2, \text{ or } 3$ . Pharmacokinetic values were  
194 analysed in GenStat version 14.0 (VSN International Ltd, Hemel Hempstead, UK) with a  
195 restricted maximum likelihood linear mixed model (REML) as mentioned above for skin  
196 temperature, limb circumference and limb sensitivity. Pair-wise comparison was undertaken  
197 in Excel (Microsoft Excel® 2016 MSO) for any significant treatment x timepoint interactions  
198 to compare differences across timepoints and treatments, using least significant differences  
199 (LSD). LSD was calculated using the formula:  $1.96 \times \text{average predicted standard error of}$   
200 differences (SED).

201 Data plotted in figures are predicted means  $\pm$  standard error of the mean (SEM), with the  
202 exception of limb sensitivity and gait score which are reported as probabilities.  $P < 0.05$  was  
203 considered statistically significant.

## 204 **Results**

### 205 ***Animals***

206 The mean  $\pm$  SD body weight of the sheep during the study phase was  $56.8 \pm 6.69$ kg. No  
207 adverse effects were observed following intramuscular or subcutaneous administration of  
208 meloxicam.

### 209 ***Skin temperature***

210 There was a significant treatment x timepoint interaction ( $p < 0.001$ ) and effect of limb on skin  
211 temperature ( $p < 0.001$ ). Ambient temperature had a significant effect on skin temperature ( $p$   
212  $< 0.001$ ). Mean temperature of the affected limb ( $23.9^\circ\text{C}$ ) was greater than control limbs  
213 ( $22.4^\circ\text{C}$ ). Skin temperature of affected limbs slightly increased over time with a marked drop  
214 in temperature observed at 6 hours post-administration, followed by a steady, continual  
215 incline until 48 hours (Table 3). There was no significant difference between treatments until  
216 12 hours after administration where treatments: S/C 1.0mg/kg and IM 2.0mg/kg were  
217 significantly less than the control (Table 3). At 24 hours, S/C 1.0mg/kg resulted in a  
218 significantly lower skin temperature than all treatments except S/C 2.0mg/kg.

### 219 ***Limb circumference***

220 There was a significant treatment x timepoint interaction ( $p = 0.012$ ) and effect of limb on  
221 limb circumference ( $p < 0.001$ ). Mean circumference of the affected limb (14.16mm) was  
222 greater than control limbs (13.74mm). Limb circumference of affected limbs increased slightly  
223 over time with an observed drop in limb circumference at 1 hour and 12 hours post drug

224 administration (Table 3). The S/C 2.0mg/kg treatment resulted in a significantly lower limb  
225 circumference than IM 2.0mg/kg at 0.5 hours after administration with no other significant  
226 differences observed between treatments until 10-12 hours post administration where the  
227 IM 1.0mg/kg treatment had a significantly lower circumference than the control treatment.

### 228 ***Limb sensitivity***

229 There was a significant treatment x timepoint interaction ( $p < 0.001$ ) and effect of limb on limb  
230 sensitivity ( $p = 0.01$ ). The maximum limb sensitivity threshold (10kg) was achieved by 65.3%  
231 of affected limbs compared to 76.4% of control limbs, indicating a greater limb sensitivity  
232 associated with affected limbs. Limb sensitivity increased over time, with all treatments  
233 reaching the maximum threshold at 0 hours compared to zero treatments at 24-48 hours  
234 post-drug administration (Table 3). Significant differences between treatments over time  
235 were observed with all treatments demonstrating reduced limb sensitivity when compared  
236 to the control at 6 hours post drug administration. At 48 hours, treatments: S/C 1.0mg/kg, IM  
237 2.0mg/kg and S/C 2.0mg/kg had significantly reduced limb sensitivity when compared to the  
238 control.

### 239 ***Gait score***

240 There was a significant treatment x timepoint interaction for gait score ( $p < 0.001$ ). Lameness  
241 was most apparent from 0.5 to 2 hours post drug administration with a decrease apparent  
242 over the 48 hours (Figure 2). A lameness score of 1, 2 or 3 was most probable between 0.5  
243 and 8 hours, with 1-hour post-administration associated with the highest probability of a  
244 lameness score of 3. At 48 hours post drug administration, the S/C 2.0mg/kg treatment was  
245 the only one observed to return gait of all sheep to a baseline score of 0. Treatments: S/C  
246 1.0mg/kg (94.6%) and IM 2.0mg/kg (82.2%) also demonstrated a high likelihood of achieving  
247 a lameness score of 0, with the IM 1.0mg/kg treatment (33.8%) less likely to achieve this score

248 than the control treatment (52.9%). Whilst there was a trend for a significant treatment  
249 effect, pair-wise comparison revealed no differences between treatments were significant.

### 250 ***Pharmacokinetic analysis***

251 The mean serum concentrations versus time for meloxicam administered IM at 1.0mg/kg or  
252 S/C at 1.0mg/kg were graphically displayed (n=3) (Figure 3). The mean C<sub>max</sub> for meloxicam  
253 administered IM 1.0mg/kg (n=3) was 10.62 ± 1.56 µg/mL and was achieved at a mean T<sub>max</sub>  
254 of 1.17 ± 0.76 h. The mean C<sub>max</sub> for meloxicam administered S/C 1.0mg/kg was 7.23 ± 0.75  
255 µg/mL and was achieved at a mean T<sub>max</sub> of 4.67 ± 1.15 h. Due to time constraints calculations  
256 of pharmacokinetic parameters of IM 1.0mg/kg and S/C 1.0mg/kg were calculated from 6  
257 sheep (Table 4). There was a significant treatment x timepoint interaction (p <0.001) for the  
258 meloxicam concentrations within plasma. The IM 1.0mg/kg treatment had a significantly  
259 higher meloxicam plasma concentration when compared to S/C 1.0mg/kg at timepoints: 0.5,  
260 1, 2 and 4 hours post drug-administration. There was no significant difference between the  
261 two treatments at any other timepoints.

### 262 **Discussion**

263 The present study was the first conducted to assess the pharmacokinetics and efficacy of  
264 meloxicam when administered intramuscularly and subcutaneously in sheep. Since  
265 pharmacokinetic data for NSAID's cannot be extrapolated between species,<sup>1</sup> these two routes  
266 of administration and alternative doses to current recommendations were investigated in  
267 order to determine whether more efficacious pain management is available. Analysis of study  
268 results indicated that meloxicam was partially efficacious at providing analgesia in sheep  
269 injected with oil of turpentine.

270 Sheep demonstrated an immediate hyper-acute response to the turpentine injection by  
271 demonstrating restricted weight bearing on the affected limb as well as lying and pawing  
272 behaviour. Agitation was evident with the affected limb being held off the ground or  
273 frequently raised, up until 12 hours post-administration, with all affected limbs  
274 demonstrating a significantly higher skin temperature, limb circumference and limb  
275 sensitivity when compared to the control limb. In several sheep, irritation persisted up until  
276 48 hours, slightly longer than the 24 hours previously observed in literature.<sup>18</sup> This method  
277 has been previously used to assess the efficacy of oral administration of NSAID's including  
278 flunixin and carprofen in sheep.<sup>19</sup>

279 Skin temperature and limb circumference were selected as measurement points as both are  
280 indicators of inflammation and presumably would emphasize the anti-inflammatory action of  
281 meloxicam on oedema and vasodilation.<sup>10,18</sup> Both were significantly affected by meloxicam  
282 administration, however proved ineffective at evaluating the efficacy of alternative  
283 treatments against the control, with zero treatment variations observed until 12 hours after  
284 drug administration. Similar observations were identified in previous studies where skin  
285 temperature and limb circumference did not significantly differ between NSAID treated ewes  
286 and those only administered turpentine, with weak to no effects observed.<sup>10,18,19</sup>

287 As ambient temperature had a large effect on skin temperature, wool present on the forelimb  
288 during measurements may have limited accurate measurement of indicators in this study as  
289 meloxicam has previously been observed to affect local skin temperature in horses at various  
290 doses.<sup>10</sup> Skin temperature increases observed from 24-48 hours in this study were likely  
291 related to a decline in meloxicam concentration in the blood as skin temperature of

292 turpentine affected sheep remained elevated when compared to the the control for up to 72  
293 hours in a previous study.<sup>18</sup>

294 Studies utilising limb circumference as an assessment of the analgesic efficacy of meloxicam  
295 vary. Alternatively, meloxicam has been found to return limb circumference to that of control  
296 limbs 4 hours earlier than sheep receiving only turpentine, yet also demonstrate no effect in  
297 other studies.<sup>10,18</sup> This study found a slight increase in limb circumference over time, similar  
298 to literature assessing efficacy of alternative NSAID's, however only the IM 1.0mg/kg  
299 treatment differed significantly from the turpentine-only sheep at 12-24 hours post  
300 administration.<sup>19</sup> This may be related to the large limb circumference measurements at  
301 baseline which were approximately 2-3mm larger when compared to Colditz et al. (2011)  
302 resulting in a less marked increase over time and difficulty in assessing efficacy of alternative  
303 meloxicam treatments.

304 Limb sensitivity has previously been effective in assessing the anti-nociceptive action of  
305 meloxicam in sheep.<sup>18</sup> Technical issues with the algometer meant that data on limb sensitivity  
306 was limited to only one phase of this study and therefore data was difficult to analyse. Data  
307 was therefore converted to a binomial score whereby any sheep achieving a threshold of  
308 10kg/f were assigned a score of 1 and those unable to achieve this threshold, assigned a score  
309 of 0. Limb sensitivity was observed to increase over time, with meloxicam treatments playing  
310 a significant role in limb desensitisation. This correlates with additional pharmacokinetic  
311 studies indicating meloxicam administered intravenously at 1.0mg/kg can alleviate pain-  
312 related signs of induced lameness.<sup>1</sup> At 6 hours post-administration, limbs were significantly  
313 less sensitive in all meloxicam treated groups when compared to the control. From 8-10  
314 hours, all treatments were significantly similar, however meloxicam treatments: S/C

315 1.0mg/kg, IM 2.0mg/kg and S/C 2.0mg/kg were again associated with decreased limb  
316 sensitivity when compared to the control treatment at 12 hours. This agrees with current  
317 literature where limb sensitivity was increased in sheep receiving no meloxicam for pain relief,  
318 confirming the anti-nociceptive effects of meloxicam previously observed.<sup>18</sup>

319 The anti-nociceptive action of meloxicam however has been disputed as analgesia was  
320 ineffective in mulesed Merino lambs administered meloxicam subcutaneously at 0.5mg/kg.<sup>20</sup>  
321 Paull et al. (2008) assessed gait using an ethogram, however studies utilising alternative  
322 methods for assessing gait, such as the numerical scoring used in this study, achieved similar  
323 results. It was observed that in the administration of alternative NSAID's including flunixin  
324 and carprofen, analgesic efficacy could not be determined,<sup>19</sup> indicating that methodology  
325 likely does not play a role in these findings. Comparisons between various studies indicate  
326 there may be a breed difference associated with pain as studies using Merino's have had  
327 limited success when using gait as a measure of efficacy. However, the study by Colditz et al.  
328 (2011) utilising Merino x Romney ewes proved somewhat successful indicating treatment  
329 with meloxicam was associated with a lower lameness score when compared to the control  
330 at 8 and 24 hours post-administration. This notion of pain disparity has been previously  
331 recognised indicating that pain responses greatly vary depending on species, sex, age, body  
332 size and even between individual animals as it is a multifaceted experience affected by a  
333 variety of physical and non-physical factors such as previous experience and actual tissue  
334 damage.<sup>19,21,22</sup>

335 Whilst trends were observed in gait score, this study found no significant differences between  
336 treatments, similar to previous studies studying Merino's. Lameness has previously been  
337 observed to peak at 24 hours post-administration of turpentine,<sup>18</sup> however this study



338 observed the opposite, with all treatments demonstrating a high likelihood towards a low  
339 lameness score when compared to the control . Pharmacokinetic analysis in this study assists  
340 in identifying the cause of the lameness peak at 24 hours previously observed in literature.  
341 The observed peak correlates to a marked decrease in meloxicam plasma concentrations  
342 between 12 to 24 hours after administration of turpentine.

343 The pharmacokinetic data obtained in this study indicate that intramuscular administration  
344 at 1.0mg/kg results in higher meloxicam plasma concentration over the first 4 hours post-  
345 administration, has a longer elimination half-life and achieves pain relief more rapidly than  
346 subcutaneous at 1.0mg/kg. Time constraints limited this study and the additional treatments:  
347 IM 2.0mg/kg, S/C 2.0mg/kg and the control were not analysed. When compared to previous  
348 studies, meloxicam plasma concentrations were higher after subcutaneous (7.23 µg/mL) and  
349 intramuscular (10.62 µg/mL) administration at 1.0mg/kg than oral (1.72 µg/mL)  
350 administration at 0.99mg/kg and intravenous administration (4.25 µg/mL) at 0.5mg/kg in  
351 sheep.<sup>1,4</sup>

352 Whilst intravenous doses in sheep have only been administered at 0.5mg/kg, literature  
353 indicates that meloxicam action is independent of dose and thus comparability is possible.<sup>1,23</sup>  
354 Two studies have previously investigated the pharmacokinetics of intravenous administration  
355 of meloxicam at 0.5kg/mg, both varying in terms of the elimination half-life achieved. Shukla  
356 et al. (2007) reported an elimination half-life of 10.85 hours, whereas Stock et al. (2013)  
357 reported a far more superior half-life of 14.0 hours, which exceeds that of both intramuscular  
358 (12.47h) and subcutaneous administration (10.24h) achieved in this study at half the dose.  
359 This disparity may once again be a result of breed and sex difference with one study examining  
360 Dorset cross males/females and the other crossbreed female sheep.<sup>1,4</sup> Comparison with

361 previous literature indicates that both subcutaneous and intramuscular administration of  
362 meloxicam are more effective than alternative methods and could be recommended for use  
363 in sheep. However, further research of pharmacokinetics in sheep may need to account for  
364 potential breed and sex differences observed across various studies.

365 Whilst improved pharmacokinetic parameters were observed for intramuscular  
366 administration when compared to subcutaneous at 1.0mg/kg, plasma concentration of  
367 meloxicam was only significantly higher in intramuscular administration over the first 4 hours  
368 of inflammation. This may suggest that whilst an extended half-life and increased maximum  
369 plasma concentrations were observed, the two treatments are equally as efficacious, with  
370 intramuscular more effective at providing short-term pain relief only. Therefore, the risks and  
371 challenges associated with on-farm administration and pain management associated with  
372 intramuscular administration of NSAID's should be considered. Intramuscular administration  
373 leads to an increase in serum creatine kinase (CK) activity resulting in potential myonecrosis  
374 and muscle damage in livestock, thus requiring precise technique for injection and on-label  
375 use in regards to dosage requirements on-farm.<sup>24-26</sup> This may prove difficult in on-farm  
376 settings where large numbers of sheep require pain management for husbandry procedures  
377 such as tail docking and castration. Further studies are required to ensure the safety of  
378 intramuscular administration of meloxicam and to determine any toxicity risks associated  
379 with double doses at 2.0mg/kg.

## 380 **Conclusion**

381 This study demonstrates that meloxicam is effective at providing some analgesia after  
382 injection of turpentine into the limb in sheep. Whilst some knowledge of analgesic efficacy  
383 according to dosage and administration route over time were obtained, this study could not

384 significantly distinguish analgesic efficacy of meloxicam between treatments in sheep. Further  
385 research is required before alternative routes and doses of meloxicam should be registered  
386 for use in sheep.

### 387 **Acknowledgements**

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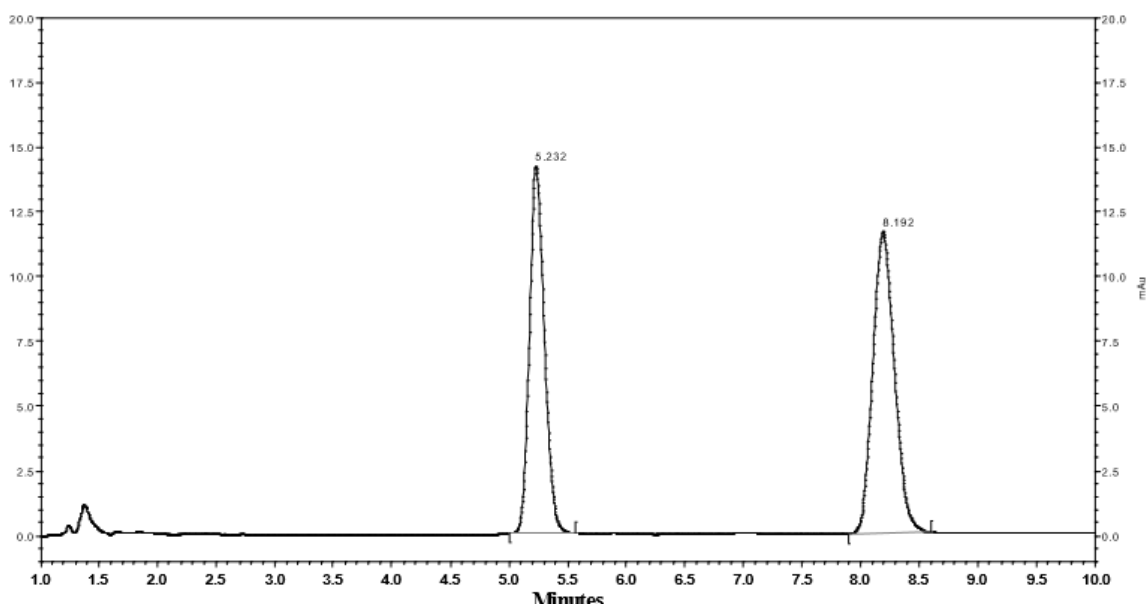
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452 **Figures**

453 **Table 1.** Randomised crossover design experimental treatment groups

Week 1			Week 2	
Group	Route of administration	Dose	Route of administration	Dose
1	Subcutaneous	1.0mg/kg	Intramuscular	2.0mg/kg
2	Intramuscular	1.0mg/kg	Subcutaneous	2.0mg/kg
3	Subcutaneous	2.0mg/kg	Control	
4	Intramuscular	2.0mg/kg	Subcutaneous	1.0mg/kg
5	Control (no meloxicam)		Intramuscular	1.0mg/kg



454 **Figure 1.** Chromatogram representing drug observation time from HPLC analysis of blank  
 455 plasma spike with meloxicam (8.5 minutes) and IS (5.5 minutes).  
 456

457 **Table 2.** NRS used to score behavioural characteristics associated with lameness in sheep  
 458 injected with oil of turpentine and meloxicam.

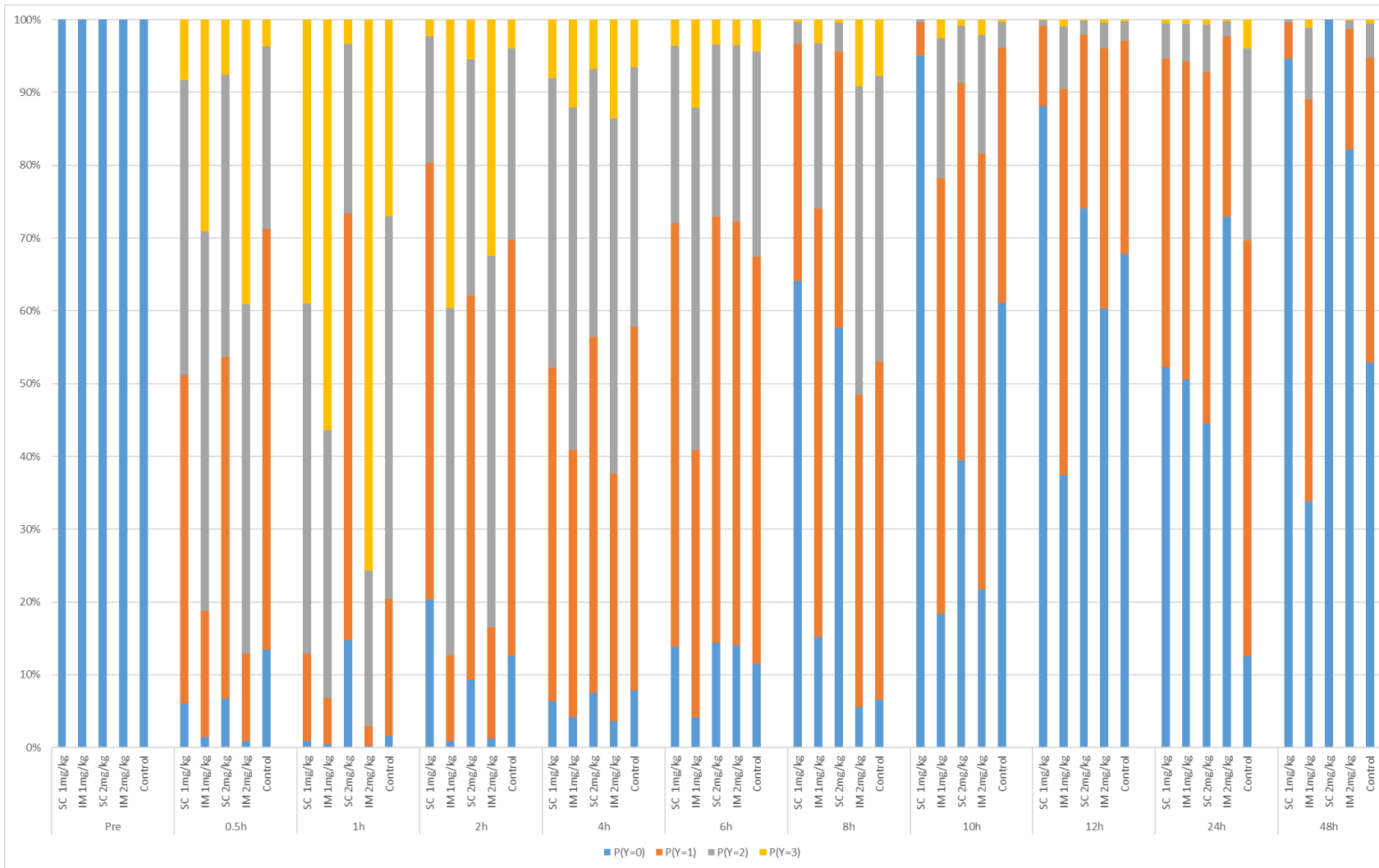
Score	Associated behaviour
0	Even distribution across all limbs with no abnormality in gait
1	Mild favouring of limbs, however all limbs used to work
2	Some limping, however all limbs used when waling with reluctance to place limb on ground
3	Severe abnormality of gait demonstrated by limited weight bearing on affected limb, increased lying behaviour and pawing affected limb

459 **Table 3.** Predicted means±SEM of skin temperature, limb circumference and limb sensitivity (1.0 = no sensitivity, 0 = highly sensitive) across  
 460 meloxicam treatment groups and timepoint.

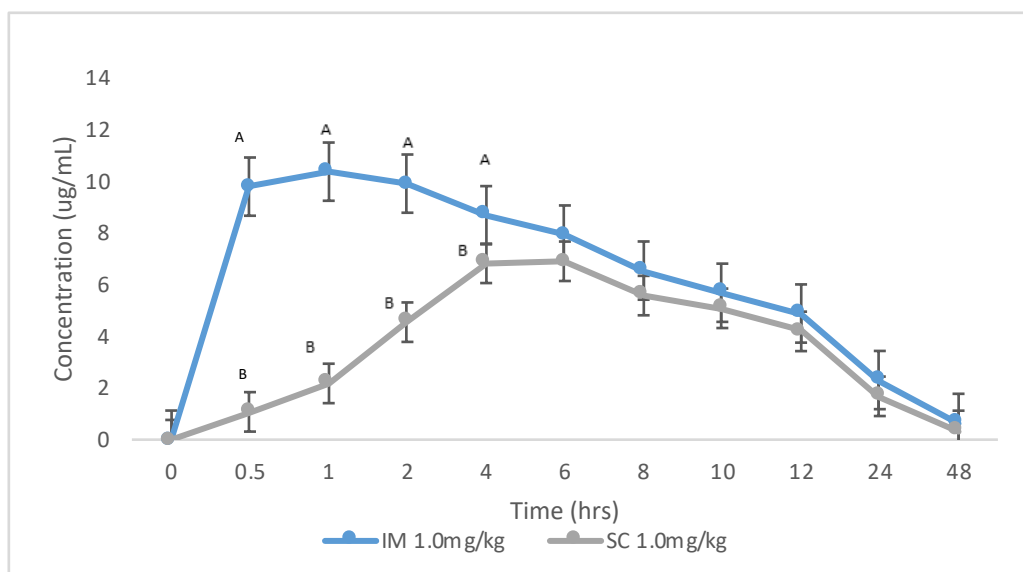
Treatment vs. Timepoint	0	0.5	1	2	4	6	8	10	12	24	48
<i>Skin temperature (°C)</i>											
<b>S/C 1.0mg/kg</b>	19.03±0.94 <sub>A</sub>	22.86±0.94 <sub>B</sub>	23.93±0.94	24.78±0.94	25.93±0.94 <sub>A</sub>	21.91±0.94 <sub>B</sub>	23.64±0.94	22.44±0.94	21.50±0.94 <sub>a</sub>	18.35±0.94 <sub>a</sub>	25.16±0.94
<b>IM 1.0mg/kg</b>	18.8±0.94 <sub>A</sub>	22.39±0.94 <sub>B</sub>	22.57±0.94	24.48±0.94 <sub>A</sub>	28.61±0.94 <sub>B</sub>	22.30±0.94 <sub>A</sub>	24.82±0.94	22.88±0.94	23.11±0.94 <sub>ab</sub>	22.25±0.94 <sub>ab</sub>	23.90±0.94
<b>SC 2.0mg/kg</b>	21.03±0.94	22.32±0.94	23.9±0.94	25.76±0.94	26.12±0.94 <sub>A</sub>	22.03±0.94 <sub>B</sub>	24.62±0.94	22.72±0.94	23.06±0.94 <sub>ab</sub>	21.05±0.94 <sub>Aab</sub>	24.67±0.94 <sub>B</sub>
<b>IM 2.0mg/kg</b>	19.08±0.94	22.32±0.94	22.16±0.94	23.55±0.94	25.50±0.94 <sub>A</sub>	21.79±0.94 <sub>B</sub>	23.07±0.94	21.94±0.94	21.81±0.94 <sub>a</sub>	22.45±0.94 <sub>b</sub>	24.59±0.94
<b>Control</b>	19.28±1.15 <sub>A</sub>	23.97±1.15 <sub>B</sub>	22.46±1.15	24.73±1.15	27.64±1.15 <sub>A</sub>	23.87±1.15 <sub>B</sub>	24.90±1.15	23.87±1.15	25.25±1.15 <sub>b</sub>	22.11±1.15 <sub>Ab</sub>	25.97±1.15 <sub>B</sub>
<i>Limb circumference (mm)</i>											
<b>S/C 1.0mg/kg</b>	13.68±0.31	14.19±0.31 <sub>ab</sub>	13.13±0.31 <sub>A</sub>	14.12±0.31 <sub>B</sub>	14.14±0.31	14.11±0.31	14.14±0.31	14.23±0.31 <sub>a</sub>	13.99±0.31 <sub>ab</sub>	13.85±0.31	14.02±0.31
<b>IM 1.0mg/kg</b>	13.53±0.31	13.78±0.31 <sub>ab</sub>	12.92±0.31	13.53±0.31	13.97±0.31	13.63±0.31	13.56±0.31	13.70±0.31 <sub>a</sub>	13.39±0.31 <sub>a</sub>	13.79±0.31	13.90±0.31
<b>SC 2.0mg/kg</b>	13.68±0.31	13.43±0.31 <sub>a</sub>	13.56±0.31	13.98±0.31	13.81±0.31	13.86±0.31	13.91±0.31	14.04±0.31 <sub>a</sub>	13.76±0.31 <sub>ab</sub>	14.12±0.31	14.04±0.31
<b>IM 2.0mg/kg</b>	14.17±0.31	14.52±0.31 <sub>b</sub>	13.74±0.31	14.09±0.31	14.08±0.31	14.12±0.31	14.34±0.31	14.28±0.31 <sub>a</sub>	14.07±0.31 <sub>ab</sub>	14.43±0.31	14.65±0.31
<b>Control</b>	13.96±0.42	13.69±0.38 <sub>ab</sub>	13.30±0.38	14.05±0.38	14.25±0.38	14.06±0.38	14.39±0.38	14.61±0.38 <sub>b</sub>	14.30±0.38 <sub>b</sub>	14.43±0.38	14.36±0.38
<i>Limb sensitivity (%)</i>											
<b>S/C 1.0mg/kg</b>	1.0±0.18	0.83±0.18 <sub>abc</sub>	0.83±0.18	1.0±0.18	0.83±0.18	0.67±0.18 <sub>ac</sub>	0.67±0.18	0.83±0.18	1.0±0.18 <sub>ac</sub>	0.67±0.18	0.83±0.18 <sub>a</sub>
<b>IM 1.0mg/kg</b>	1.0±0.18	1.0±0.18 <sub>ad</sub>	1.0±0.18	1.0±0.18	0.67±0.18	0.5±0.18 <sub>a</sub>	0.5±0.18	0.67±0.18 <sub>A</sub>	0.17±0.18 <sub>Bb</sub>	0.5±0.18	0.17±0.18 <sub>c</sub>
<b>SC 2.0mg/kg</b>	1.0±0.18 <sub>A</sub>	0.5±0.18 <sub>Bab</sub>	1.0±0.18 <sub>A</sub>	0.83±0.18	0.67±0.18	1.0±0.18 <sub>c</sub>	0.83±0.18	0.5±0.18	0.83±0.18 <sub>ac</sub>	0.5±0.18	0.83±0.18 <sub>a</sub>
<b>IM 2.0mg/kg</b>	1.0±0.18 <sub>A</sub>	0.5±0.18 <sub>Bab</sub>	0.83±0.18	0.83±0.18	0.67±0.18	0.67±0.18 <sub>ac</sub>	0.5±0.18	0.67±0.18	0.5±0.18 <sub>ab</sub>	0.67±0.18	0.5±0.18 <sub>ac</sub>
<b>Control</b>	1.0±0.22	1.0±0.22 <sub>c</sub>	1.0±0.22 <sub>A</sub>	1.0±0.22 <sub>B</sub>	0.5±0.22 <sub>A</sub>	0±0.22 <sub>b</sub>	1.0±0.22	0.5±0.22	1.0±0.22 <sub>Ac</sub>	0±0.22 <sub>B</sub>	0±0.22 <sub>Bc</sub>

461 \*Means in a column without a common subscript (a, b, c) represent significant differences between treatments whilst means in a row without  
 462 a common subscript represent significant differences between timepoints (A, B) (p <0.005).





463 **Figure 2.** Probabilities of lameness score (Y = 0, 1, 2 or 3) when administered subcutaneously and intramuscularly at 1.0mg/kg  
 464 over time.



465 **Figure 3.** Concentration of meloxicam when administered subcutaneously and  
 466 intramuscularly at 1.0mg/kg over time. Data points represent predicted mean  $\pm$  SEM for n =  
 467 3 per group. \*Subscripts (A,B) represent significant differences between treatments  
 468 (p < 0.05).

469 **Table 4.** Mean  $\pm$  SD pharmacokinetic parameters of 1.0mg/kg meloxicam when administered  
 470 intramuscularly and subcutaneously in sheep.

	IM 1.0mg/kg	S/C 1.0mg/kg
<b>Tmax (hrs)</b>	1.17 $\pm$ 0.76	4.67 $\pm$ 1.15
<b>Cmax (<math>\mu</math>g/mL)</b>	10.62 $\pm$ 1.56	7.23 $\pm$ 0.75
<b>AUC<sub>0-t</sub> (<math>\mu</math>g/h/mL<sup>-1</sup>)</b>	168.95	121.76
<b>t<sub>1/2</sub> (hrs)</b>	12.47	10.24
<b>kel</b>	0.0556	0.0677

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