

Frequencies of Morphological Sperm Abnormalities in Australian Bulls and Rams

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23 *1. Abstract*

24 Levels of abnormal sperm within an ejaculate can influence the reproductive productivity
25 in livestock. This study aimed to establish sperm abnormality frequencies as per the Australian
26 Cattle Veterinarians eight-category system amongst Australian bulls and rams. It was
27 hypothesised rams would present lower levels of sperm abnormalities than bulls based on the
28 higher normal sperm sheep industry standard. Semen samples were sourced from bulls (n =
29 300) and rams (n = 149) of various breeds located throughout NSW and SA and were fixed in
30 5% formalin in buffered saline and stored in a 4°C before assessment. Differential interference
31 contrast optics at x400 magnification were used to count 100 cells per sample, which were
32 categorised as per the Australian Cattle Veterinarians eight-category system. Both rams and
33 bulls followed similar trends in frequencies across all eight sperm morphology categories with
34 loose heads/abnormal tails being the most prevalent (11.54% and 10.23% respectively).
35 Percentage mean of normal sperm was significantly higher in bulls (74.03%) when compared
36 to rams (68.05%), rejecting the hypothesis ($P < 0.01$). The findings in this paper allows a
37 preliminary understanding of what can be expected to appear in the average Australian ram
38 ejaculate. Further investigation into abnormality frequencies as characterised by the eight-
39 category system within a larger ram population would gain understanding of normal levels of
40 sperm abnormalities within a ram ejaculate.

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42 *Key words*

43 Bulls, frequencies, rams, sperm abnormality, sperm evaluation, sperm morphology

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46 2. *Introduction*

47 The prevalence of abnormal sperm within an ejaculate can influence the sample quality and
48 fertility of the male. In comparison to cattle, morphological sperm assessments in rams are
49 seldom performed alongside breeding soundness evaluations. Knowing average frequencies of
50 sperm abnormalities provides clinicians with insight into the level of normal and abnormal
51 sperm likely to be seen in a population of bulls or rams.

52 Acceptable thresholds of outlined sperm abnormalities in livestock determines whether
53 individuals pass or fail breeding soundness exams based on sperm morphology. Thresholds for
54 normal levels of abnormal sperm in bulls is well established amongst literature. Fordyce et al.
55 (2006) outlines proximal cytoplasmic droplets, pyriform heads and knobbed acrosomes
56 abnormalities have an acceptable threshold of 20%. Midpiece abnormalities, loose
57 heads/abnormal tails, vacuoles and teratoids and swollen acrosomes have an acceptable
58 threshold of 30% within bull ejaculates (Fordyce et al., 2006). The threshold determined in
59 rams is more simply, no more than 15% of abnormal sperm present in an ejaculate (S.P. de
60 Graaf, personal communication). In rams, morphological standards are not set to the same
61 standard creating an ambiguous threshold of acceptable levels of abnormal sperm and their
62 links to fertility. Little is known about the representation of various types of sperm
63 abnormalities within the ejaculate of ram semen, with current data characterising ejaculate
64 makeups of small sample sizes ($n \leq 20$; Almadaly et al., 2016; Azubuike et al., 2017; Basioura
65 et al., 2022). Furthermore, industry standards of normal levels of abnormal sperm and their
66 relationship to ram fertility are yet to be determined. Frequencies of abnormalities will vary
67 depending on how sperm are categorised.

68 Detailed classification systems, such as the eight-category system used with Australian bulls
69 (Fordyce et al., 2006; Koziol & Armstrong, 2018), provide reproductive biologists with insight

70 into which specific abnormality is impacting the percentage of normal sperm. As certain
71 abnormalities have been linked to external stressors and sire reproductive performance, this
72 knowledge can inform the identification of sub-fertile males (Barth, 1986; Söderquist et al.,
73 1991). When assessing ram semen, sperm are simply classified as normal or abnormal (Evans
74 & Maxwell, 1987). Without characterising subcategorization of the abnormal category, type
75 and subsequent frequency of abnormalities remains unknown. Without detailed categorisation
76 of abnormalities, it hinders insight into the root causes of these abnormalities as well as their
77 potential relationship with negative fertility.

78 By having detailed categorisation of abnormalities, identifying the presence of particular
79 abnormalities can pinpoint influences on the fertility of individual animals. Of all possible
80 abnormalities, head morphometry has demonstrated the closest link to livestock fertility in
81 current literature (Barth et al., 1992). This has been stated as early as 1927 by Williams and
82 Savage (Saacke, 2004). In rams, sperm subpopulations based on head dimensions have been
83 correlated to fertility rate (Bravo et al., 2014; Maroto-Morales et al., 2015; Santolaria et al.,
84 2015). Furthermore, sperm with detached heads or tailless sperm are considered immobile
85 because they can't migrate through the female tract. Bulls with over 70% of detached heads or
86 tailless sperm are considered to have decreased fertility (Perry, 2021). Knowing this link
87 between abnormalities and fertility should drive future research to further understanding causes
88 of sperm abnormalities. This can provide more insight into why individuals not meeting
89 breeding soundness thresholds set by the cattle or sheep industries.

90 This study aimed to establish frequencies of sperm abnormalities in ram and bull ejaculates
91 from properties across New South Wales (NSW) and South Australia (SA), Australia, using the
92 eight-category system. By characterising ram ejaculates in populations over 20 individuals this
93 study addresses the gap in the literature to provide a preliminary understanding of the expected
94 percentage of normal and abnormal sperm within Australian ram ejaculates and compare this

95 to bull ejaculates. Industry thresholds for accepted abnormalities within ram ejaculates is 15%
96 (S.P. de Graaf, personal communication), while in cattle this is doubled at 30% (Perry, 2021).
97 Based on these thresholds, it is hypothesised there will be lower frequencies of sperm
98 abnormalities present amongst rams when compared to bulls.

99 *3. Materials and methods*

100 *3.1. Morphological assessment of sperm*

101 Samples from rams (n = 149) and bulls (n = 300) were sourced from 16 Australian
102 properties across the states of New South Wales and South Australia. They were fixed in 5%
103 formalin in buffered saline and stored in a 4°C walk in freezer prior to analysis. Samples were
104 vortexed to ensure homogeneity prior to assessment, then 9µl of each sample was pipetted onto
105 a glass slide and covered with a 22x22mm coverslip. As an observational study, samples were
106 observed under an Olympus BX53 microscope with Differential Inference Contrast (DIC)
107 optics at 400x magnification to count 100 sperm per sample. Overlapping or partially focused
108 sperm were excluded from counts to avoid incorrect categorisation.

109 Sperm were categorised as per the ACV eight-category system into normal sperm, proximal
110 cytoplasmic droplets, midpiece abnormalities, loose heads and abnormal tails, pyriform heads,
111 knobbed acrosomes, vacuoles and teratoids and swollen acrosomes (each category detailed
112 further in Perry (2021)). This category system was chosen because it is used in the ACV
113 standardised BullCheck Scheme in assessing bull fertility for breeding (BullCheck Scheme,
114 2024). Frequencies for individual sires and categories were recorded in Microsoft Excel
115 (version 2407).

116 *3.2. Methods of accuracy/reliability*

117 To ensure observer accuracy prior to data collection training samples were used to
118 ensure consistency with expert morphologists. Prior to commencing assessment, the observer
119 was required to reliably record results within $\pm 10\%$ of the expert morphologists' results.
120 Accuracy of counting was ensured by using an automatic counter which alerted the user when
121 100 sperm were recorded. Sessions were broken up with multiple breaks to reduce fatigue and
122 ensure reliability of the observer. In cases of sperm with multiple abnormalities, abnormalities
123 closer to the head took precedence, due to the close link between head abnormalities and
124 fertility (Sprecher & Coe, 1996).

125 *3.3. Minimising bias*

126 Observer bias was managed in the categorisation of sperm using descriptions of
127 normality/abnormality provided in Perry (2021), and comparison of visual aids as per Seymour
128 et al. (2023).

129 *3.4. Data analysis*

130 To ensure confidentiality, data identifiers were removed including owner, property
131 name and sire ID/name and replaced with generic numeric identifiers. In Microsoft Excel
132 (version 2407) descriptive statistics of each morphological category were calculated, as well
133 as figures obtained using Pivot Charts and Pivot Tables. Standard one tailed t-tests were
134 conducted to make interspecies comparisons of normal levels of sperm in ejaculates.

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139 4. Results

140 4.1. Mean percentages of sperm abnormality categories were similar for bulls and rams

141 The mean percentage of each category in both rams and bulls was compared (Figure 1).
142 There was a similar trend in average across all eight categories in both rams and bull. The
143 highest mean value recorded was loose heads/abnormal tails for both bulls and rams (10.23%
144 and 11.54% respectively). Midpiece abnormalities (7.65% in bulls, 10.83% in rams) and
145 proximal cytoplasmic droplets (5.69% in bulls, 7.95% in rams) were found next most prevalent
146 in ejaculates. Amongst midpiece and loose heads/abnormal tail abnormalities, bulls displayed
147 more extreme outliers in comparison to rams. The lowest recorded mean value of any
148 abnormality was swollen acrosomes in both rams and bulls (0% and 0.03% respectively).

149 4.2. Percentage of mean normal sperm was significantly higher in bulls in comparison to
150 rams

151 Variations were present in mean percentage of normal sperm (PNS) found within the
152 ejaculates of each species (Figure 2A). The mean of normal sperm was significantly higher (P
153 < 0.01) in bull ejaculates (74.03%) in comparison to ram ejaculates (68.05%). Interestingly, the
154 standard deviation was higher in rams (SD = 25.83) than bulls (SD = 18.98). PNS in the bull
155 population had a minimum of 5% and maximum of 99% (Figure 2B). Similarly, rams had a
156 minimum PNS of 10% and maximum of 100% (Figure 2C).

157 4.3. Bulls passed the cattle industry threshold of $>70\%$ percentage normal sperm
158 significantly more than rams

159 Finally, the proportion of individuals that passed cattle ($>70\%$) and sheep ($>85\%$) industry
160 normality standards was quantified for this population (Figure 3). The number of bulls that
161 passed the cattle industry threshold was significantly higher than rams ($P < 0.001$). When the

162 threshold was >85%, only one third of ejaculates from each species pass the sheep industry
163 standard threshold.

164 5. *Discussion*

165 This study aimed to establish frequencies of sperm abnormalities in Australian ram and bull
166 ejaculates from 16 properties across NSW and SA, Australia. Abnormalities were categorised
167 as per the complex eight-category system used in bulls. Frequencies of abnormalities in all
168 categories followed a similar trend when comparing bulls and rams. Industry thresholds outline
169 normal sperm levels need to be higher than <70% for cattle and <85% for sheep ejaculates, and
170 hence hypothesised lower levels of sperm abnormalities would be present within rams than
171 bulls. The findings from this study reject this hypothesis.

172 Frequencies of sperm abnormalities were comparably similar of bulls and rams across all eight
173 categories. Mean percentage of abnormalities followed similar trend in both rams and bulls
174 (Figure 1). Loose heads/abnormal tails were the most prevalent (10.23% in bulls, 11.54% in
175 rams), while swollen acrosomes were non-existent to rare (0.03% in bulls, 0% in rams) in
176 ejaculates of both species. Loose heads and abnormal tails found in this study were much higher
177 than previous literature. In bulls this abnormality is found in frequencies <6% (Azubuike et al.,
178 2017; Callaghan et al., 2016; Menon et al., 2011a; Söderquist et al., 1991). More limited in
179 rams, the small number of findings available showed these frequencies do not exceed 5%
180 within an ejaculate which is contrast to the 10.23% observed in the population of rams in this
181 study (Azubuike et al., 2017; Savage, 1984; Yániz et al., 2012). More similarly to our finding,
182 15.25% of abnormal tails were observed within a ram ejaculate, however it was identified as
183 having varicocele (Basioura et al., 2022).

184 Swollen acrosomes were non-existent in our ram population and rare amongst bulls (0.03%,
185 Figure 1). These were consistent with ram findings of no distorted or twisted acrosomes the

186 study of López Armengol et al. (2018). They were similarly identified in bulls at 0.1%
187 (Callaghan et al., 2016), as well as present within 6% of bulls in the work of Andersson et al.
188 (1989), however the frequency of this abnormality is not mentioned. Acrosomal swelling is
189 considered normal after sperm undergo the acrosome reaction, suggesting swollen acrosomes
190 present after premature offset of the acrosome reaction (Perry, 2021). The low prevalence of
191 swollen acrosomes found in both species could be relevant to the levels of knobbed acrosomes
192 abnormality within an ejaculate (Perry, 2010), of which were also found in low prevalence in
193 this study. Considering the location of properties being in NSW and SA, animals are at a higher
194 risk of experiencing heat stress during warmer months. This may explain the high frequency
195 of abnormal tails and proximal cytoplasmic droplet abnormalities (Barth & Bowman, 1994;
196 Perry, 2021) shown in this study, but would need to be confirmed with future studies.

197 The mean percentage of normal sperm (PNS) present within an ejaculate was varied between
198 rams and bulls (Figure 2). Significantly higher levels of normal sperm were present in bulls
199 (74.03%) than rams (68.05%), meaning more abnormalities were present in ram (Figure 2A),
200 rejecting the hypothesis. PNS values of other bull studies ranged from 76% - 92.4%, only
201 slightly higher (< 2%) than what was observed in this population (Al-Saedi & Abdulkareem,
202 2022; Callaghan et al., 2016; Freneau et al., 2010; Menon et al., 2011a; Menon et al., 2011b;
203 Walters et al., 2004). PNS values of rams were notably higher amongst other studies, ranging
204 from 82.25% - 96.58% (Basioura et al., 2022; Hassan et al., 1970; Yániz et al., 2012). The
205 difference in PNS values between literature could be attributed to the different definitions of
206 normality considered from each paper. Within this study normal bull and ram sperm were
207 characterised as those with an intact acrosome, a smooth, rounded paddle shaped head and a
208 long tail that lacks twists, breaks, coils or bends. Furthermore, abaxial tails, bent midpieces
209 (bent to at least 90° without folding or breaking), segmental aplasia, distal cytoplasmic droplets
210 slightly pyriform heads and narrow heads were also considered normal due to their lack of

211 influence on fertility, as per Perry (2021). Koziol and Armstrong (2018) also characterises distal
212 cytoplasmic droplets as normal sperm in bull assessments. Given the differences in PNS values
213 of this study and literature in both species, more notably sheep, the characterisation of
214 normality and abnormality should be stated in future sperm morphology studies.

215 Interestingly, rams showed more variability in the standard deviation of mean PNS than bulls
216 (SD = 25.83 in rams, SD = 18.98 in bulls). This finding indicates bulls may have a higher
217 susceptibility to internal or external stressors in comparison to rams. Stressors could include
218 environmental, health-related or handling practises, including semen collection (Fernandez-
219 Novo, 2020; Palmer, 2005). Current literature shows there are various reasons for abnormalities
220 presenting within the ejaculate. Heat stress has been linked to being the cause of multiple
221 abnormalities including loose/detached heads, abnormal tails, proximal cytoplasmic droplets
222 (Basioura et al., 2022; López Armengol et al., 2018; Perry, 2021). Variation could also stem
223 from the number of properties ram ejaculates were sourced from when compared to bull
224 ejaculates (11 and 5 respectively), despite the sample size of rams being almost halved. Two
225 ram studies noted a PNS value of 5%, where in both cases rams were affected by varicocele
226 (Ott et al., 1982; Savage, 1984), which more similarly relates to the minimum PNS found in
227 rams was 10% (Figure 2C). It would be interesting to explore whether this was due to disease
228 like varicocele or another factor in future. The higher variability found in ram ejaculates could
229 be attributed to lower susceptibility to stressors or number of properties ejaculates were sources
230 from in comparison to bulls.

231 Using different industry threshold for acceptable levels of normal sperm within an ejaculate
232 altered the number of individuals that passed these thresholds. Significantly more bulls than
233 rams passed the cattle industry standard of >70% PNS in an ejaculate (Figure 3). Interestingly,
234 when increased to the sheep industry standard of >85%, only one third of either species passed
235 the threshold. Industry thresholds of PNS are used as an indicator for fertility success in

236 breeding soundness exams (Bravo et al., 2014; Söderquist et al., 1991). The bull industry
237 threshold of >70% PNS (Koziol & Armstrong, 2017) is more feasible in practice because there
238 will be more sires included in the breeding program. Using the sheep industry standard of
239 >85%, may not be appropriate for this population due to the low pass rate of ~30% of bulls or
240 rams.

241 Whilst the ram population size (n = 149) in this study is the largest of any previous literature
242 found (n ≤ 20), it is recognised the sample size may not provide an accurate representation of
243 the total NSW, SA or Australian ram population. The findings of this study do provide a
244 preliminary insight into the makeup of the average ejaculate of NSW and SA rams and the
245 frequencies of normal and abnormal sperm in samples. and how this can be applied to fertility
246 data already done in large scale studies performed in bulls. There is literature available
247 comparing the effectiveness of sperm evaluation methodologies in cattle. This is not the case
248 currently in rams and is suggested to be performed in future to see whether using these cattle-
249 based protocols is truly appropriate for use in sheep studies. Building upon the population size
250 in future research would be an interesting comparison to these findings and may gain further
251 insight into the makeup of normal ram ejaculates. Lastly, a limitation of manual sperm
252 assessments relies on the characterisation of abnormalities by the observer, where in this study
253 was only a single morphologist, increasing the potential for bias in this study. Bias of the
254 morphologist was minimised through cross referencing training samples with expert
255 morphologist data, as well as closely following written and visual references (Perry, 2021;
256 Seymour et al., 2023).

257 This study has significance in being the first study to focus on the frequency of sperm
258 abnormalities in the ram in a population over 20 individuals, to provide a preliminary insight
259 into the makeup of a ram ejaculate based on a detailed sperm categorisation system. It was
260 hypothesised rams would have lower levels of sperm abnormalities in an ejaculate in

261 comparison to bulls. Bulls had a significantly higher PNS than rams, rejecting the hypothesis.
262 Bulls and rams follow similar patterns of normality and abnormality frequencies, with loose
263 heads/abnormal tails, midpiece and proximal cytoplasmic droplets being most prevalent among
264 both species. Similarly, swollen acrosomes were the least prevalent in both species. Levels of
265 abnormalities amongst larger populations of rams and bulls should be explored to further the
266 findings of this study in the future. The information presented will be useful for breeders and
267 clinicians by allowing them to categorise sperm abnormalities more accurately and understand
268 what could be expected to appear in an average ejaculate.

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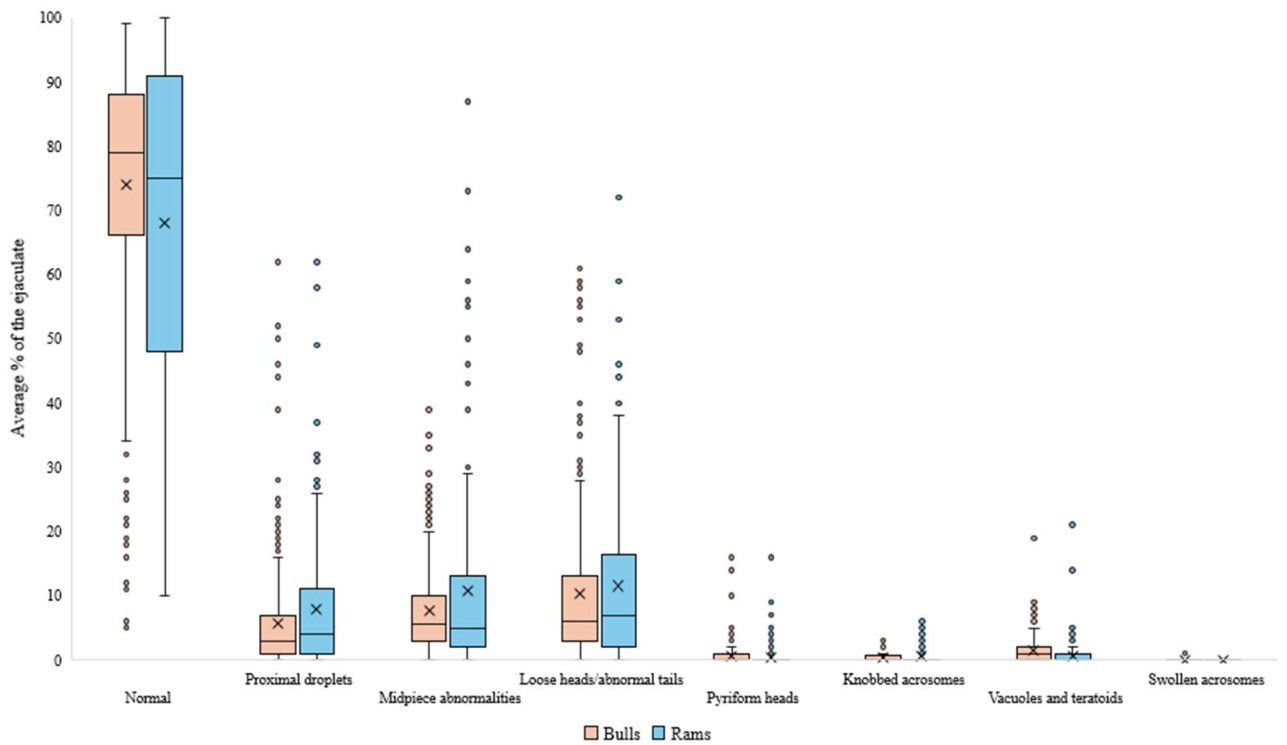
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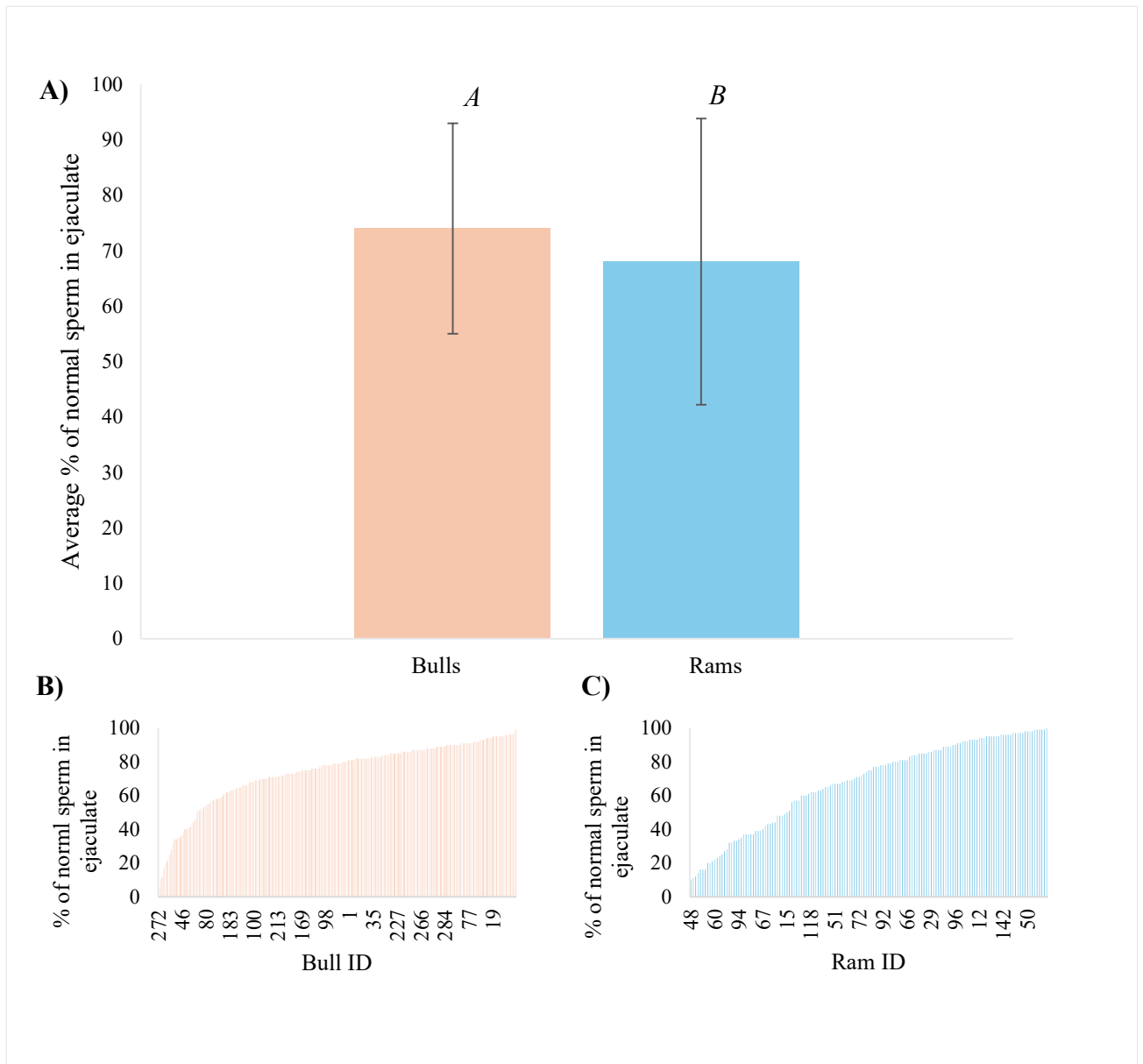


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284 *Figure 1: Sperm morphology characteristics of bull (n = 300) and ram (n = 149) ejaculates*
 285 *classified using an eight-category system (1. Normal sperm, 2. Midpiece abnormalities, 3.*
 286 *Proximal cytoplasmic droplets, 4. Loose heads/abnormal tails, 5. Pyriform heads, 6. Knobbed*
 287 *acrosomes, 7. Vacuoles and teratoids, 8. Swollen acrosomes).*

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293 *Figure 2: A) Variation in percentage of normal sperm (PNS) in bull (n = 300, SD = 25.83) and*

294 *ram (n = 149, SD = 18.98) ejaculates. Values without common superscripts differ significantly*

295 *(P < 0.001). B) Range of normal sperm amongst bull ejaculates (n = 300, min = 99%, max =*

296 *5%). C) Range of normal sperm amongst ram ejaculates (n = 149, min = 100%, max = 10%).*

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300 *Table 1: Proportion of ejaculates in bulls (n = 300) and rams (n = 149) that pass respective*
301 *cattle and sheep industry thresholds of normal sperm within an ejaculate to satisfy soundness*
302 *breeding evaluations. Values without common superscripts differ significantly (P < 0.001).*

	Cattle industry standard (>70% PNS)	Sheep industry standard (>85% PNS)
Bulls	69.3% (208/300) ^A	32% (96/300)
Rams	53.7% (80/149) ^B	33.6% (50/149)

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